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HEPATOPROTECTIVE ROLE OF *YUCCA GLORIOSA L.* EXTRACT AGAINST CCl₄ INDUCED HEPATOTOXICITY IN RATS

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ABSTRACT

Yucca gloriosa L. commonly known as Spanish Dagger and Family- Agavaceae. It is a stemless or rising of stature of small trees and trunk short. The whole plant of *Yucca gloriosa L.* is used for ulcer, jaundice, asthma and bronchitis. To investigate the hepatoprotective activity and acute oral toxicity of extract of whole plant of *Yucca gloriosa L.* (PYG) in male Wistar albino rats by using CCl₄ induced hepatotoxicity. The PYG at doses of 200 and 400mg/kg, p.o and the standard drug Silymarin (100mg/kg, p.o) were administered three times at 12h intervals and then CCl₄ (1ml/kg) was administered to all the groups except normal control for 2 days. The hepatoprotective activity was assessed by using various biochemical parameters like SGOT, SGPT, ALP, γ -GT, TP and total bilirubin along with histopathological studies were observed after 36h of CCl₄ treatment. The PYG at the doses of 200 and 400mg/kg inhibited CCl₄ induced liver toxicity in Wistar albino rats as assessed by the biochemical changes and histopathological studies. The pet ether extract of whole plant of *Yucca gloriosa L.* afforded significant protection against CCl₄ induced hepatocellular injury.

Key words: *Yucca gloriosa L.*, Hepatoprotective, CCl₄, Silymarin, Hepatotoxicity.

INTRODUCTION

The liver is the largest organ in the body weighing 1200-1500g. It is a key organ in regulating homeostasis within the body. It regulates several important functions including protein synthesis, storage and metabolism of fats and carbohydrates, detoxification of drugs and other toxins, metabolism of hormones and excretion of bilirubin. Liver diseases are associated with distortion of these metabolic functions [1,2]. Although viruses are the main cause of liver diseases, the liver lesions arising from xenobiotics, excessive drug therapy, environmental pollution and alcoholic intoxication are not uncommon [3]. Every year about 20,000 deaths are found due to liver disorders [4]. Thus to maintain a healthy liver is a crucial factor for

overall health and well beings [5]. Thus, liver diseases remain one of the serious health problems and its disorders are numerous with no effective remedies [6-8]. There is no rational therapy available for treating liver disorders and management of liver diseases is still a challenge to the modern medicine [9-11]. In the absence of reliable liver protective drugs in allopathic medical practices, herbs play a role in the management of various liver disorders [6]. The use of natural remedies for the treatment of various hepatic diseases has a long history and medicinal plants and their derivatives are still used all over the world [4].

Yucca gloriosa L. commonly known as Spanish Dagger and Family- Agavaceae. It is a stemless or rising of stature of small trees and trunk short. This plant leaves are 2-3 feet long, 2 in wide, long pointed, often tooth margin, mostly in rosettes at surface of ground or ends of trunk. Flowers has many cup or saucer shaped, hanging, greenish white to reddish, fragrant, born mostly in erect panicles that usually overtop the leaves. The whole plant of

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Yucca gloriosa L. is used for ulcer, jaundice, asthma and bronchitis [12]. Therefore we attempt to investigate the hepatoprotective activity of this plant against CCl₄-induced liver damage in rats to support the claim. Hence, the present study was designed to verify the claims of the native practitioners.

MATERIALS AND METHODS

Plant material

The whole plant of *Yucca gloriosa L.* was collected from Tirumala hills, Tirupati, Andhra Pradesh, India. It was identified and authenticated by Prof. Madhava Chetty, K., Taxonomist, S.V. University, Tirupati, Andhra Pradesh, India. A voucher specimen has been kept in our laboratory for future reference.

Preparation of plant extract

The collected whole plant was dried at room temperature, pulverized by a mechanical grinder, sieved through 40mesh. About 100g of powdered materials were extracted with petroleum ether (60°-80°C) using soxhlet apparatus. The extraction was carried out until the extractive becomes colourless. The extracts is then concentrated and dried under reduced pressure. The solvent free semisolid mass thus obtained is dissolved in tween 80 and used for the experiment. The percentage yield of prepared extract was around 10.5%w/w.

Animals Used

Albino rats (180–200 g) of either sex were maintained in a 12 h light/dark cycle at a constant temperature 25 °C with free access to feed (Sai durga feeds and foods, Bangalore) and water. All animals were fasted prior to all assays and were allocated to different experimental groups each of 6 rats. Moreover the animals were kept in specially constructed cages to prevent coprophagia during the experiment. All experiments were carried out according to the guidelines for care and use of experimental animals and approved by Committee for the Purpose of Control and Supervision of Experiments on Animals (CPCSEA). Ethical committee clearance was obtained from IAEC (Institutional Animal Ethics Committee) of CPCSEA.

Acute toxicity study

The procedure was followed according to the OECD guidelines 423 (Acute toxic class method). The acute toxic class method is a step wise procedure with 3 animals of single sex per group. Depending on the mortality and or moribund status of the animals, on an average 2-4 steps may be necessary to allow judgment on the acute toxicity of the testing substance. According to this procedure minimum number of animals were to be used for acceptable data band scientific conclusion. The method uses defined doses (5, 50, 300, 2000 mg/kg body weight) and the results allow a substance to be ranked and classified

according to the globally harmonized system (GHS) for the classification of chemical which causes acute toxicity.

Adult female wistar rats were used for this study. The starting dose of whole plant of *Yucca gloriosa L.* extract was 2000 mg/kg body weight, as most of the crude extracts possess LD₅₀ value more than 2000 mg/kg body weight. The dose was administered to overnight fasted rats and food was withheld for a further 3-4 hours after administration of the drug and observed for signs of toxicity.

Body weight of the rats before and after treatment were noted and any changes in skin, eye, and mucous membranes, salivation, nasal discharge, urination and behavioral (sedation, depression), neuromuscular (tremors, convulsions), cardiovascular, lethargy, sleep and coma were noted. The onset of toxicity was also noted. The animals were kept under observation for 14 days.

The acute toxicity of Petroleum Ether extract of *Yucca gloriosa L.* whole plant was determined as per the OECD guideline no. 423 (Acute Toxic Class Method). It was observed that the test extract was not lethal to the rats even at 2000mg/kg dose. Hence, 1/10th (200mg/kg) and 1/5th (400mg/kg) of this dose were selected for further study [13].

Carbon tetrachloride induced hepatotoxicity in rats

The liver protective effect was evaluated using the carbon tetrachloride (CCl₄) model described by *Rao and Mishra* [14]. Wistar albino rats (150-200g) were divided into five groups and were subjected to the following treatments; group-I served as normal control; received vehicle only. Group-II served as untreated group; received only CCl₄, to assist assessing the severity of toxicity produced by carbon tetrachloride administration. Groups III-V served as treated groups; received PYG at the dose of 200 and 400mg/kg, p.o. and standard drug Silymarin at a dose of 100mg/kg, p.o. were administered orally to rats of the respective groups three times at 12h intervals. Carbon tetrachloride diluted with liquid paraffin (1:1) was administered in dose of 1ml/kg, p.o. for 2 days to all animal groups except for normal control. After 36h of carbon tetrachloride treatment, blood was collected from all groups of rats by puncturing the retro-orbital sinus. Serum was separated by centrifugation at 2500rpm at 37^oC for 15min and analyzed for various biochemical parameters.

Biochemical estimation

The separated serum was subjected to estimate SGOT and SGPT by *Reitman and Frankel* method [15], alkaline phosphatase (ALP) by *Kind and King* method [16], and bilirubin by *Malloy and Evelyn* method [17].

Statistical analysis

The data were expressed as mean ± standard error

mean (S.E.M).The Significance of differences among the group was assessed using one way and multiple way analysis of variance (ANOVA). The test followed by Tukey-Kramer multiple comparison tests, the p values less than 0.05 were considered as significance.

RESULTS

Acute toxicity study

The body weight of the rats before and after administrations were noted that there is slightly increased the body weight. But there are no changes in skin and fur, eyes, mucous membranes, respiratory, circulatory, autonomic and central nervous system and motor activity and behavior pattern were observed and also no sign of tremors, convulsions, salivation, diarrhea, lethargy, sleep and coma were noted. The onset of toxicity and signs of toxicity also not there. In this study there was no toxicity/death were observed at the dose of 2000mg/kg b.w. The acute toxicity study in rats showed that at 2000 mg/kg dose, the plant is safe for consumption and for medicinal uses (Table 1). In the acute toxicity study, the animals treated with the PYG at a higher dose of 2000 mg/kg did not

manifest any significant abnormal signs, behavioral changes, body weight changes, or macroscopic findings at any time of observation. There was no mortality in the above-mentioned dose at the end of the 14 days of observation.

Effect of PYG on CCl₄ – induced hepatotoxicity

The results of PYG on Carbon tetrachloride-induced hepatotoxicity were represented in Table 2. The animals treated only with CCl₄ exhibited a significant increase ($P<0.001$) the levels of SGOT, SGPT, ALP, γ -GT and total bilirubin as well as decrease in the levels of TP when compared to the normal control group after 36h of CCl₄ treatment, indicating hepatocellular damage. The PYG at tested doses (group-III & IV) produced a significant reduction ($P<0.001$) in the CCl₄ induced elevated levels of SGOT, SGPT, ALP, γ -GT and total bilirubin as well as increases the TP when compared to the animals treated only with CCl₄ (group-II) after 36h of CCl₄ treatment. Overall, PYG at tested doses significantly reduced the levels of hepatic enzymes and total bilirubin.

Table 1. Acute toxicity study of pet ether extract of *Yucca gloriosa* L. Linn. (PYG) in rats

S.No	Groups	Dose/kg b.w, p.o	Weight of animals		Signs of Toxicity	Onset of Toxicity	Duration of study
			Before Test	After Test			
1	PYG	2000 mg	175 g	180 g	No signs of Toxicity	Nil	14days
2	PYG	2000 mg	195 g	205 g	No signs of Toxicity	Nil	14days
3	PYG	2000 mg	190g	195 g	No signs of Toxicity	Nil	14days
4	PYG	2000 mg	195 g	200 g	No signs of Toxicity	Nil	14days
5	PYG	2000 mg	215 g	220 g	No signs of Toxicity	Nil	14days
6	PYG	2000 mg	195 g	205 g	No signs of Toxicity	Nil	14days

Table 2. Effects of PYG on alternation of hepatic enzyme and serum bilirubin in rat after 36h. of CCl₄ treatment

Groups (n=6)	Biochemical Parameters					
	SGOT (U/L)	SGPT (U/L)	ALP (U/L)	γ -GT (IU/L)	TP (gm/dl)	Total Bilurubin (mg/dl)
Group-I (Normal Control)	35.64 \pm 0.67***	23.43 \pm 0.34***	192.42 \pm 0.18***	53.48 \pm 1.16***	9.55 \pm 0.18***	0.87 \pm 0.04***
Group-II (CCl ₄ : 1ml/kg)	68.49 \pm 1.19	40.53 \pm 0.34	426.22 \pm 0.69	104.45 \pm 0.41	2.43 \pm 0.12	3.74 \pm 0.04
Group-III (PYG: 200mg/kg)	48.19 \pm 0.22***	33.57 \pm 0.46***	254.39 \pm 0.19***	62.84 \pm 0.34***	3.47 \pm 0.27***	1.24 \pm 0.04***
Group-IV (PYG: 400mg/kg)	42.5 \pm 0.12***	25.47 \pm 0.26***	226.59 \pm 0.48***	59.37 \pm 0.22***	5.49 \pm 0.43***	0.84 \pm 0.02***
Group-V (Silymarin: 100mg/kg)	35.12 \pm 0.17***	21.68 \pm 0.46***	199.73 \pm 0.68***	52.47 \pm 0.43***	7.65 \pm 0.54***	0.74 \pm 0.03***

Values are expressed as mean \pm SEM of 6 rats in each group. *** $p<0.001$, as compared to CCl₄-treated group. SGOT = Serum glutamate oxaloacetate transaminase, SGPT = Serum glutamate pyruvate tranaminase, ALP = Alkaline phosphatase, γ -GT = Gamma glutamyl transpeptidase, TP = Total proteins.

DISCUSSION AND CONCLUSION

Liver is the vital organ of metabolism and excretion. It produces and secretes bile; it also produces fibrinogen, prothrombin, heparin and sulfuric acid ester. The liver converts sugar into glycogen [18]. Any changes in anatomy or functions of liver are characterized by cirrhosis, jaundice, tumors, liver cell necrosis and hepatitis, metabolic and degenerative lesion etc. The management of hepatic diseases is still a challenge to the modern medicines [10,19]. Herbal medicines play a major role in the treatment of liver disorders. A number of medicinal plants and their formulations are widely used for the treatment of these disorders [20,21]. However, there were not enough scientific investigations on the hepatoprotective activities conferred to these plants. One of the plants from Indian flora is *Yucca gloriosa L.* The present studies were performed to investigate the hepatoprotective activity of pet ether extract of whole plant *Yucca gloriosa L.* in rats against carbon tetrachloride as hepatotoxin to prove its claims in folklore practice against liver diseases.

Carbon tetrachloride (CCl_4) is one of the most commonly used hepatotoxins in the experimental study of liver diseases [22]. CCl_4 is potent hepatotoxin producing centrilobular hepatic necrosis. It is accumulated in hepatic parenchyma cells and metabolized to trichloromethyl free radicals (CCl_3) by liver cytochrome P-450 dependent monooxygenases. This CCl_3 free radical combined with cellular lipids and proteins in the presence of oxygen to produce lipid peroxides [23]. Thus, antioxidant or free radical generation inhibition is important in protection against CCl_4 induced liver lesion [24]. The flavonoids constituents possess free radical scavenging properties [25].

In general, the extent of liver damage is assessed by histopathological evaluation and levels of hepatic enzymes such as ALP, SGOT, SGPT and also Bilirubin release in circulation [26,27]. The estimation of gamma glutamyl transpeptidase (γ -GT) is an important screening test with a high negative predictive value for hepatic disease [28].

Administration of hepatotoxins CCl_4 elevated the serum levels of SGOT, SGPT, ALP, γ -GT and bilirubin as well as decreases total serum proteins (TP) significantly [29,30]. The rise in serum enzymes level and bilirubin has been attributed to the damaged structural integrity of the liver, because they are cytoplasmic in location and released into circulation after cellular damages [31].

In our investigation, the biochemical changes were observed after 36h of CCl_4 treatment. Thereby, it was found that the animal groups which are pretreated with PYG at the

dose of 200 and 400mg/kg (groups-III and IV) as well as silymarin at the dose of 100mg/kg (group-V) for three times at 12h. intervals, resulted in significantly decreases the hepatic enzymes such as SGOT, SGPT, ALP and γ -GT and also total bilirubin; as well as increases the total serum proteins (TP) as compared to animals treated only with CCl_4 (group-II). These results give us the suggestion that, the animals which are pretreated with PYG as well as silymarin, showed a protection against the injurious effects of CCl_4 that may results from the interference with cytochrome P-450. These biochemical restoration may be due to the inhibitory effects on cytochrome P-450 or/and promotion of its glucuronidation [32,33]. Silymarin is a known hepatoprotective drug. It is reported to have a protective effect on the plasma membrane of hepatocytes [34].

In histopathological assessment, it was found that the normal liver architecture was disturbed by CCl_4 intoxication. In the liver section of rats treated with PYG showed the ability of PYG to prevent hepatocellular necrosis, thereby further confirming the significant hepatoprotective effect of whole plant of *Yucca gloriosa L.*

It is well documented that the phytoconstituents comes under the category of flavonoids, alkaloids, glycosides, carotenoids, phenols, coumarins, lignans, essential oil, lipids, monoterpenes, xanthenes and organic acids are reported to have hepatoprotective activity [35]. Literature review revealed that various chemical investigations were carried out with this plant. *William Carey Mamidipalli et al.*, have been reported the preliminary phytochemical screening of the pet ether extract of *Yucca gloriosa L.* revealed that presence of steroids, flavonoids, tannins, alkaloids and glycosides [36]. *Mulabagal vanisree et al.*, have been reported that the purification of the pet ether extract yielded n-hentriacontane, ferulic acid, 4-hydroxycinnamic acid, quercetin-3-rhamnoside and kaempherol-3-glucoside; along with beta-sitosterol, beta-sitosterol-glucoside and d-manitol [37]. The hepatoprotective activity of *Yucca gloriosa L.* may be attributed due to presence of these constituents. This study supports the traditional claims and the PYG could be added in traditional preparations for the various liver diseases.

It is concluded from the data, that the pet ether extract of whole plant of *Yucca gloriosa L.* possesses significant hepatoprotective activity and may prove to be effective for the treatment of liver disorders. However, longer duration studies on chronic models are necessary to elucidate the exact mechanism of action so as to develop it as a potent hepatoprotective drug.

REFERENCES

1. Ward FM, Daly MJ. Liver disease. In: Roger walker, Clive Edwards. Churchill Livingstone, New York, *Clinical Pharmacy and Therapeutics.*, 3, 2005, 209.

2. Wolf PL. Biochemical diagnosis of liver disease. *Indian Journal of Clinical Biochemistry.*, 14, 1999, 59–90.
3. Baskar Rajan G, Chezhiyan N. Strength and Wealth of Therapeutic medicinal plants iIndia,In: Irfan A Khan, Atiya Khanum, Role of Biotechnology in Medicinal and Aromatic plants, Special volume on Diseases. *Ukaaz publications Hyderabad.*, 6, 2002, 151-152.
4. Lewis HW, Elvin-Lewis MPH. Plants affecting man's Health, John Wiley and Sons, New York. *Medical Botany.*, 1977, 217-218.
5. Sharma A, Chakraborti KK, Handa SS. Anti-hepatotoxic activity of some Indian herbal formulations as compared to Silymarin. *Fitoterapia.*, 62, 1991, 229-235.
6. Handa SS, Sharma A, Chakraborti KK. Natural products and plants as liver protecting drugs. *Fitoterapia.* 57, 1986, 307-45.
7. Venkateswaran S, Pari L, Viswanathan P, Menon VP. Protective effect of Livex; a herbal formulation against erythromycin estolate induced hepatotoxicity in rats. *Journal of Ethnopharmacol.*, 57 (3), 1997, 161–167.
8. Latha U, Rajesh MG, Latha MS. Hepatoprotective effect of an ayurvedic medicine. *Indian Drugs.* 36, 1999, 470–473.
9. Chandrasekar VM, Abdul Haseeb TS, Nagappa AN. Hepatoprotective activity of *Wrightia tinctoria* in rats. *Indian Drugs.* 41, 2004, 366.
10. Meyer SA, Kulkarni AP. Hepatotoxicity, In: Hodgson E, and Smart RC, Introduction to biochemical toxicology. John Wiley and Sons, New York. *Medical Botany.*, 3, 2001, 487-490.
11. Surendar Angothu, Mohana Lakshmi S, Saravana Kumar A, Yalla Reddy K. epatoprotective activity of Antigonon leptopus Hook. & Arn. against carbon tetrachloride (CCl₄) induced hepatotoxicity in Wistar albino rats. *International Journal of Biological & Pharmaceutical Research*, 1(1), 2010, 27- 32.
12. Madhava chetty K. et al. "*Yucca gloriosa L.*" Flowering Plants of Chittoor District, Andhra Pradesh, India. 2008.
13. OECD, 2002. Acute oral toxicity. Acute oral toxic class method guideline 423 adopted 23.03.1996, In: Eleventh Addendum to the, OECD, guidelines for the testing of chemicals organisation for economical co-operation and development, Paris, June, 2000
14. Rao KS, Mishra SH. Anti-inflammatory and hepatoprotective activities of fruits of *Moringa pterygosperma gaertn.* *Indian Journal of natural Products.*, 1998, 14: 3.
15. Reitman S, Frankel S. A colorimetric method for the determination of SGPT and SGOT. *American Journal of Clinical Pathology.*, 28, 1957, 56-62.
16. Kind PRN, King EJ. Determination of Serum Alkaline Phosphatase. *Journal of Clinical Pathology.* 7, 1954, 132-136.
17. Malloy HT, Evelyn KA. The determination of bilirubin with the photoelectric colorimeter. *Journal of Biological Chemistry.*, 119 (2), 1937, 481-485.
18. Nadeem MPC, Dandiya PC, KV, Pasha M, Imran D, Balani K, Vohora SB. Hepatoprotective activity of *Solanum nigrum* fruits. *Fitoterapia.*, 68, 1997, 245-251.
19. Harsh Mohan. Text book of pathology. *Jaypee Publisher.*, 4, 2002, 569-630.
20. Subramonium A, Puspangadan P. Development of Phytomedicines for liver diseases. *Indian journal of Pharmacology.*, 31, 1999, 166-175.
21. Thyagarajan SP, Jayaram S, Gopalakrishnan V, Hari R, Jayakumar P, Sripathi MS. Herbal medicines for liver diseases in India. *J Gastroenterol Hepatol.*, 17, 2002, 370-376.
22. Recknagel RO, Glende EA, Dolak JA, Walter RL. Mechanism of carbon tetrachloride toxicity. *Pharmacology and Therapeutics.*, 43, 1989, 139-154.
23. Johnson DE, Kroening C. Mechanism of early carbon tetrachloride toxicity in cultured rat hepatocyte. *Pharmacol toxicol.*, 83, 1998, 231-239.
24. Suja SR, Latha PG, Pushpangadan P, Rajasekharan S. Evaluation of hepatoprotective effects of *Helminthostochys Zeylanica* (L) Hook against carbon tetrachloride-induced liver damage in Wistar rats. *Journal of Ethnopharmacol.*, 92, 20, 61-66.
25. Hesham R, El-Seedi, Shgeru N. Chemistry of Bioflavonoids. *Indian J Pharm Edu.*, 39, 2007, 172.
26. Plaa G, Charbonneau M. Detection and evaluation of chemically induced liver injury, In: Hayes AW. *Principals and methods of Toxicology.*, Raven press, New York, 1994, 841-846
27. Portmann B, Talbot IC, Day DW, Davidson AR. Histopathological changes in the liver following a paracetamol over dose; Correlation with clinical and Biochemical parameter. *J Pathol.*, 117, 1975, 169-180.
28. Nemesanszky E. Enzyme testing hepatobiliary disease, In: Donald W Moss, Sidney B rosarki, enzyme test in diagnosis, *Oxford University Press.*, New York, 1996, 25-59.
29. Singh B, Saxena AK, Chandan BK, Suri OP, Suri KA, Sathi NK. Hepatoprotective activity of *Verbenalin* on experimental liver damage in rodents. *Fitoterapia.*, 60, 1998, 135.
30. Kim NK, Vasmineh WG, Frejar EF, Goldman AI, Theologides A. Value of alkaline phosphatase, 5'-nucleotidase, γ -glutamyl transferase and glutamate dehydrogenase activity measurements (Single and Combined) in serum in diagnosis of metastases to the liver. *Cli chem.*, 23, 1977, 2034-2038.
31. Sallie R, Tredger JM, William R. Drug and liver. *Biopharmaceutical Drug Disposition.* 12, 1991, 251-259.
32. Wesley GC, Brater CC, Alice RJ. *Cloth's medical pharmacology.*, Mosby year Book, US, 41, 1992.

33. Gilman AG, Rall TW, Nies AS, Taxlor P. *The pharmacological basis of Therapeutics.*, Mc Graw Hill International Edition, London, (13), 1992.
34. Ramellini G, Meldoles J. Liver protection by silymarin. In vitro effect on dissociated rat hepatocytes. *Arzneimforsch. Drug Research.*, 26, 1976, 89-73.
35. Sharma SK, Ali M, Gupta J. Plants having Hepatoprotective activity. *Phytochemistry and pharmacology.*, 2, 2002, 253-270.
36. William Carey Mamidipalli, Venkata Rao Nimmagadda, Ravi Kumar Bobbala, KrishnaMohan Gottumukkala. Preliminary studies of Analgesic and Anti-inflammatory Properties of *Antigonon leptopus* Hook et Arn Roots in Experimental Models. *Journal of Health Science.*, 54 (3), 2008, 281-286.
37. Mulabagal Vanisree, Ruby L Alexander-Lindo, David L DeWitt, Muraleedharan G Nair. Functional food components of *Antigonon leptopus* tea. *Journal of Food Chemistry.*, 106, 2008, 487-492.