



International Journal of
Experimental Pharmacology

www.ijepjournal.com

**EVALUATION OF ANTI-HYPERLIPIDEMIC ACTIVITY OF
SELECTED PLANTS WITH INDIVIDUAL AND POLYHERBAL
EXTRACTION**

Srinivasa Rao Konijeti ^{*1}, G. Vidya Sagar², Sri Leela Movva³

¹Vignan Institute of Pharmaceutical Sciences, Deshmukhi, Nalgonda district, Telangana, India-508284; Department of Biotechnology, Acharya Nagarjuna University, Guntur, Andhra Pradesh, India-522510

²Srinivasarao College of Pharmacy, PM Palem, Visakhapatnam, Andhra Pradesh, India- 500041.

³Netaji Institute of Pharmaceutical Sciences, Nalgonda, Telangana, India-508252.

ABSTRACT

The present study aimed that evaluation anti hyperlipidemic activity of root and leaf of the *Bauhinia rufescens Lam*, leaves of *Cassia auriculata L*, leaves and flowers of *Mimosa pudica L* by Maceration method. Individually and combined plants extracted poly herbal extraction and screened for phytochemical study performed Preliminary Phytochemical analysis of crude extracts for root and leaf of the *Bauhinia rufescens Lam*, leaves of *Cassia auriculata L*, leaves and flowers of *Mimosa pudica L* and poly herbal extraction for Anti-Hyperlipidemic activity by Triton X 100 Induced Hyperlipidemia model, High Fat Diet (FD) induced hyperlipidemic model, Estimation of Serum total cholesterol (TC) CHOD- PAP, Estimation of serum triglycerides, Estimation of HDL-cholesterol, Estimation of LDL cholesterol, Phytochemical investigation reveals the presence of alkaloids, flavanoids, saponins, tannins, steroids, triterpenoids, carbohydrates and glycosides in poly herbal methanolic extraction and individual plant extraction, In acute toxicity studies no mortality was observed with either of the extracts even at the dose level of 2000mg/kg body weight. In the present study, the methanol extracts of three plants reduced the cholesterol and triglycerides in a manner similar to the reduction facilitated by atorvastatin. The hypolipidemic activities of atorvastatin and the methanol extract of individual and polyherbal extraction were evident in both synthesis and excretory phases of triton-induced hyperlipidemia in rats.

Keywords: *Bauhinia rufescens Lam*, *Cassia auriculata L*, *Mimosa pudica L*.

INTRODUCTION

Hyperlipidemia is a major cause of atherosclerosis and the atherosclerosis-associated conditions, such as coronary heart disease, ischemic cerebrovascular disease and peripheral vascular disease. Although the incidence of the atherosclerosis related events has declined in the united states, these condition still accounts for the majority of morbidity and mortality among middle aged and older adults, the incidence and absolute number of annual events will increase over the next decade because of epidemic of

obesity and ageing of the U.S. population [1,2]. Dyslipidemia, including hyperlipidemia and hypercholesterolemia and low level of high density of lipoproteins cholesterol HDL are major cause of increased atherogenic risk; both genetic disorders and lifestyle diet high in calories, saturated fat, and cholesterol contribute to dyslipidemia seen in developed countries around the world [3]. Severe hypertriglyceridemia (i.e. Triglyceride level of >1000mg/dl) requires therapy to prevent pancreatitis [4]. Moderately elevated triglyceride level 150 to 400mg/dl also are concern because they often occur as part of the metabolic syndrome, which includes insulin resistance, obesity, hypertension, low HDL level and substantially increased CHD risk [5,6]. Medicinal plant based drug has

Corresponding Author

Srinivasa Rao Konijeti

Email id: srinivaskonijeti@yahoo.com

now advantageous over modern drugs. As such are long history of use and better patient tolerance as well as public acceptance, renewable source cultivation and processing environmental friendly, local availability, plant may major source of lead generation. Several recent break through are gugalipid, taxol, artimesinin [7]. Medicinal plant contains so many chemical compounds which are the major source of therapeutic agents to cure human disease [8].

Bauhinia rufescens Lam. (*Fabaceae*) is a shrub usually 1-3 m high and sometimes reaching 8 m. The bark is ash-grey, smooth and very fibrous. The leaves are very small with greenish-yellow to white and pale pink flowers. Fruits aggregated with 4-10 seeds each [9] *Cassia auriculata* L. (*Fabaceae*) is a much branched shrub with smooth cinnamon brown bark and closely pubescent branchlets. The leaves are alternate, stipulate, paripinnate compound, very numerous, closely placed. Its flowers are irregular, bisexual, bright yellow and large. The fruit is a short legume, oblong, obtuse, tipped with long style base, flat, thin, papery, undulately crimped, pilose, pale brown [10]. *Mimosa pudica* L. (*Fabaceae*) it is a prickly perennial herb that grows up between 0.5- 0.9m high; leaves are bipinnate, opposite, compound in nature and sensitive to touch; flowers are axillary positioned, clustered in fluffy balls, radially symmetrical and campanulate; fruits occur in aggregate of 2-8 pods [11].

MATERIALS AND METHODS

Collection of plant and authentication

Root and leaf of the *Bauhinia rufescens* Lam, leaves of *Cassia auriculata* L, leaves and flowers of *Mimosa pudica* L was procured from Madhavachetti botanical garden, Thirupathi and was authenticated by Dr. K. Madhavachetti, Assistant Professor in Department of Botany at Sri Venkateswara University, Tirupathi.

Extraction by Simple Maceration

The root and leaf of the *Bauhinia rufescens* Lam, leaves of *Cassia auriculata* L, leaves and flowers of *Mimosa pudica* L individually and poly herbal of three plant materials are made into powder and then gone for the Maceration with sufficient quantity of methanol for 7 days. During maceration, it was shaken twice daily. On 7th day it was filtered and the filtrate was concentrated. The remaining solvent was evaporated by heating on a water bath (50°C) to get methanolic extract and the extract was stored in desiccator.

Preliminary Phytochemical Screening

The crude methanolic extract of root and leaf of the *Bauhinia rufescens* Lam, leaves of *Cassia auriculata* L, leaves and flowers of *Mimosa pudica* L individually and poly herbal extraction were tested for its different chemical groups such as alkaloids, flavanoids, tannins, steroids, saponins, fixed oils, gums and mucilages, tri-terpenoids, carbohydrates and glycosides, phytosterols.

Experimental Animals

Wister rats of either sex weighing between 180-250g. Then the animals were acclimatized for 7 days under standard husbandry conditions. Room temperature $26 \pm 2^{\circ}$ C, Relative humidity 45-55%, Light/ dark cycle - 12: 12hr, all animal studies were performed as per the guidelines of CPCSEA and Institutional Animal Ethical Committee (IAEC). CPCSEA Approval Number: VIP/GVS/ Ph.D./ IAEC/2016-17/15.

Anti hyperlipidemic study

Triton X 100 Induced Hyperlipidemia model:

Triton X 100 (TR) induced hyperlipidemic model forty two Wistar rats were randomly divided into 7 groups of 6 each. The first group was given standard pellet diet, water and orally administered with 5% CMC. The II, III, IV, V, VI, VII group animals were injected i.p. with 10% aqueous solution of Triton 400mg/kg body weight. After 72hours of triton injection, the second group received a daily dose of 5% CMC (p.o) for 7 days. The third group was administered daily dose of MEBR 400mg/kg, fourth group was administered a daily dose of MECA 400mg/kg and Group V and Group VI was administered daily dose of MEMP 400mg/kg and 400 mg/kg suspended in 5% CMC, p.o., for 7 days, after inducing hyperlipidemia. Seventh group was administered with the standard Atorvastatin 10mg/kg, p.o. for 7 days. Food was withdrawn 10hrs prior to the blood sampling. The control group animals received the vehicle in the same volume orally.

Group 1: Administered vehicle and served as normal control.

Group 2: Administered Triton X 100 (TR) and served as hyperlipidemic control.

Group 3: Administered (MEBR) Methanolic extraction of *Bauhinia rufescens* Lam (400mg/kg), p.o.

Group 4: Administered (MECA) Methanolic extraction of *Cassia auriculata* L (400mg/kg), p.o.

Group 5: Administered (MEMP) Methanolic extraction of *Mimosa pudica* L (400mg/kg), p.o.

Group 6: Administered (PHME) Polyherbal methanolic extraction (400mg/kg), p.o.

Group 7: Administered Atorvastatin (10mg/kg), p.o.

On the 8th day, blood was collected by retro-orbital sinus puncture, under mild ether anaesthesia. The collected samples were centrifuged for 15minutes at 2500rpm. Then serum samples were collected and analyzed for serum Total Cholesterol, Triglycerides, High Density Lipoprotein Cholesterol, Low Density Lipoprotein Cholesterol and Very Density Lipoprotein Cholesterol.

High Fat Diet (FD) induced hyperlipidemic model

Preparation of Feed

Normal animal food pellets were crushed in mortar and pestle to crush into small pieces and then grinded into fine powder in mixer grinder. The other ingredients i.e. cholesterol 2%, Cholic acid 1%, sucrose 40%, and coconut

oil 10% were added in the mixer grinder in an ascending order of their quantity and mixed well. This dried powder was then mixed with same quantity of water every time to make small balls of feed and later this was stored in self sealing plastic covers in refrigerator at 2°C to 8°C. The feed for normal group was prepared similarly by grinding only the normal food pellets and then mixing with water without the other excipients. This preparation of feed was done once in three days for all the animals. Fourty two Wistar rats were randomly divided into 7 groups of six each. The chronic experimental hyperlipidemia was produced by feeding the above prepared food for 21days. The rats are then given test plant extracts and Atorvastatin (10mg/kg, p.o) once daily in the morning orally for 14 consecutive days. During these days, all the groups also received fat diet in the same dose as given earlier. The hyperlipidemic control i.e., group II animals received the hyperlipidemic diet and the vehicle. The control group animals received the normal laboratory diet and vehicle. The first group was given standard pellet diet, water and orally administered with 5% CMC. The II, III, IV, V, VI, VII group animals were injected i.p. with 10% aqueous solution of Triton 400mg /kg body weight. After 72hours of triton injection, the second group received a daily dose of 5% CMC (p.o). The third group was administered daily dose of MEBR 400mg/kg, fourth group was administered a daily dose of MECA 400mg/kg and Group V and Group VI was administered daily dose of MEMP 400mg/kg and PHME 400 mg/kg suspended in 5% CMC, p.o., for 7 days, after inducing hyperlipidemia. Seventh group was administered with the standard Atorvastatin 10mg/kg, p.o. for once daily in the morning orally for 14 consecutive days. During these days, all the groups also received fat diet in the same dose as given earlier.

Group 1: Administered vehicle and served as normal control.

Group 2: Administered Triton X 100 (TR) and served as hyperlipidemic control.

Group 3: Administered MEBR (400mg/kg), p.o. and fed with FD

Group 4: Administered MECA (400mg/kg), p.o. and fed with FD

Group 5: Administered MEMP (400mg/kg), p.o. and fed with FD

Group 6: Administered PHME (400mg/kg), p.o. and fed with FD

Group 7: Administered Atorvastatin (10mg/kg), p.o. and fed with FD

On day 15, animals were anaesthetized with Diethyl ether and blood was collected by retro- orbital puncture. The blood was subjected to centrifugation for 15min at 2500rpm to obtain serum. The collected serum was analyzed for serum Total Cholesterol, Triglycerides, High Density Lipoprotein Cholesterol, Low Density Lipoprotein Cholesterol and Very Low Density Lipoprotein Cholesterol.

Biochemical estimations

On the 8th day, blood was collected by retro-orbital sinus puncture, under mild ether anaesthesia in both the experimental models. The collected samples were centrifuged for 15minutes at 2500rpm. Then serum samples were collected and analysed for serum Total Cholesterol (TC), Triglycerides (TG), High Density Lipoprotein Cholesterol (HDL-C), Low Density Lipoprotein Cholesterol (LDL-C) and Very Low Density Lipoprotein Cholesterol (VLDL-C) serum blood glucose and atherogenic index (AI).

Estimation of Serum total cholesterol (TC) CHOD- PAP

This method was used for the estimation of serum cholesterol. In this method the following were pipetted into the reaction vessel using a micro pipette. Test samples (T): 0.02ml serum, 2.00ml reaction solution; the standard sample (S): 0.02ml standard and 2.00ml reaction solution, while for the blank sample (B): 0.02ml DW and 2.00ml reaction solution. The mixture was mixed well and incubated for 10 minutes at 20 to 25°C or 5 minutes at 37°C. The absorbance was read at 505/670 nm against the reagent blank [12].

Estimation of serum triglycerides (TG)

GPO-PAP method was used to estimate the serum triglycerides. For this 0.01 ml of serum was taken in a test tube (T) in which 1ml reaction solution was added. In another test tube (S) 0.01ml standard and 1ml reaction solution were added. The solution was mixed well and incubated at 20 to 25°C for 10min. The absorbance of standard and test against reagent blank was read at 505 (500-540 nm) [13].

Estimation of HDL-cholesterol

CHOD-PAP method was used to estimate the serum HDL cholesterol level. CHOD-PAP method (Henry, 1974) was used to estimate the serum HDL cholesterol level. For this 2 ml serum was taken in a test tube and 0.5 ml of precipitation reagent was added. The mixture was shaken thoroughly and left to stand for 10min at 15 to 25°C and then centrifuged for 15min at 4000rpm. Within 2hr after centrifugation, the clear supernatant was used for the determination of HDL-C. One ml of the supernatant was taken in a test tube (T) and 1ml of reaction solution was added to it. In another test tube 0.1 ml DW was taken and 1ml reaction solution (B) was added. The mixtures were mixed thoroughly, incubated for 10min at 15-25°C or for 5min at 37°C and measured the absorbance of the sample against reagent blank at 546nm [14].

Estimation of LDL cholesterol

LDL cholesterol was estimated by using Friedwald's (1972) formula as follows:

$$\text{LDL in mg \%} = \frac{\text{Total cholesterol} - \text{HDL-C} - \text{Triglyceride}}{\dots\dots\dots}$$

Estimation of VLDL cholesterol

VLDL cholesterol was estimated by using following formula

$$\text{VLDL in mg \%} = \frac{\text{Triglyceride}}{5}$$

Hyperlipidemic rats

Phytochemical analysis of the plant extract showed different phytoconstituents viz. glycosides, phytosterols, triterpinoids, alkaloids and flavonoids. Several phytoconstituents like glycosides, triterpinoids, Saponins, alkaloids and flavonoids are known screening of anti hyperlipidemic agents. Triton physically alters very low density lipoprotein cholesterol rendering them refractive to the action of lipolytic enzymes of blood and tissues, preventing or delaying their removal from blood and tissues. Hence the antihyperlipidemic effect of MEBR, MECA, MEMP and PHME could be due to an increased catabolism of cholesterol into bile acids. Administration of PHME (400mg/kg, p.o) for 14 days in fat diet model, successfully prevented the elevation of serum Total Cholesterol, Triglycerides, Low Density Lipoproteins Cholesterol (LDL-C), Very Low Density Lipoproteins Cholesterol (VLDL-C) , and decrease of serum High Density Lipoprotein Cholesterol (HDL-C) in Fat diet model rats respectively. It has been well established that nutrition plays an important role in the etiology of hyperlipidemia and atherosclerosis. Fat diet model is used as a chronic model for induction of hyperlipidemia. In this study we chose fat diet which contains the common ingredients in our daily food. Diet containing saturated fatty acids increases the activity of HMG CoA reductase, the rate

determining enzyme in cholesterol biosynthesis; this may be due to higher availability of acetyl CoA, which stimulated the cholesterologenesis rate. Moreover, this could be associated with a down regulation in LDL receptors by the cholesterol and saturated fatty acids in the diet, which could also explain the elevation of serum LDL-C levels either by changing hepatic LDLR (LDLreceptor) activity, the LDL-C production rate or both. LCAT enzyme is involved in the transesterification of cholesterol, the maturation of HDL-C and the flux of cholesterol from cell membranes into HDL. The activity of the enzyme tends to decrease in diet-induced hypercholesterolemia. The possible mechanism of RNM may involve increase of HDL-C, which is attributed to the mobilization of cholesterol from peripheral cells to the liver by the action of Lecithin Cholesterol O-acyltransferase (LCAT) .The increased HDL-C facilitates the transport of TG or cholesterol from serum to liver by a pathway termed ‘reverse cholesterol transport’ where it is catabolised and excreted out of the body. Antihyperlipidemic activity was observed with Atorvastatin (10mg/kg p.o.), and the PHME (400mg/kg) showed better activity than MEBR, MECA, MEMP and (400mg/kg), to have anti-hyperlipidemic properties. Treatment with PHME (400mg/kg, p.o.) for 7 days successfully prevented the elevation of serum Total Cholesterol, Triglycerides, Low Density Lipoproteins Cholesterol (LDL-C), Very Low Density Lipoproteins Cholesterol (VLDL-C) , and decrease of serum High Density Lipoprotein Cholesterol (HDL-C) in Triton model rats respectively. Results are shown in Table 1 and 2 and figure 1-10.

RESULTS AND DISCUSSION

Table 1. Effect of MEBR, MECA, MEMP and PHME on serum lipid parameter levels in Triton induced Hyperlipidemic rats.

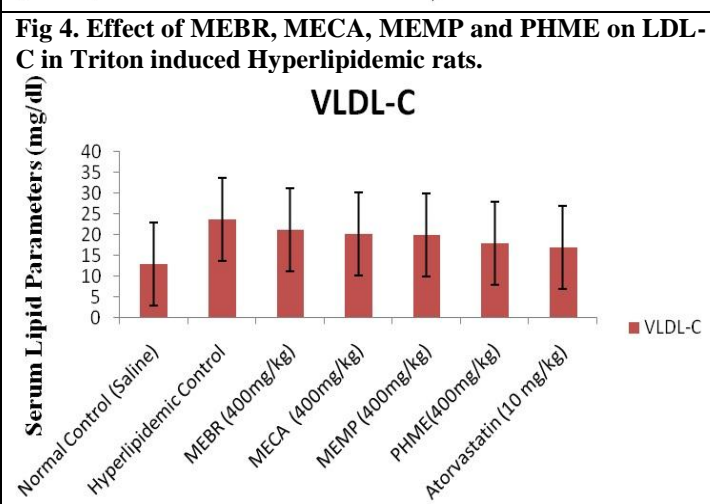
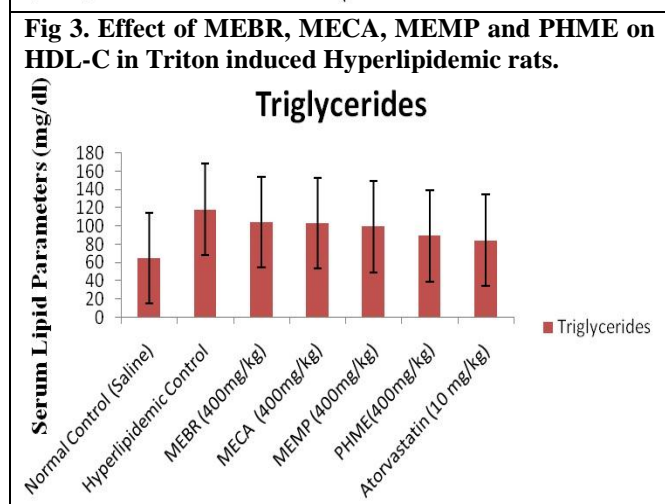
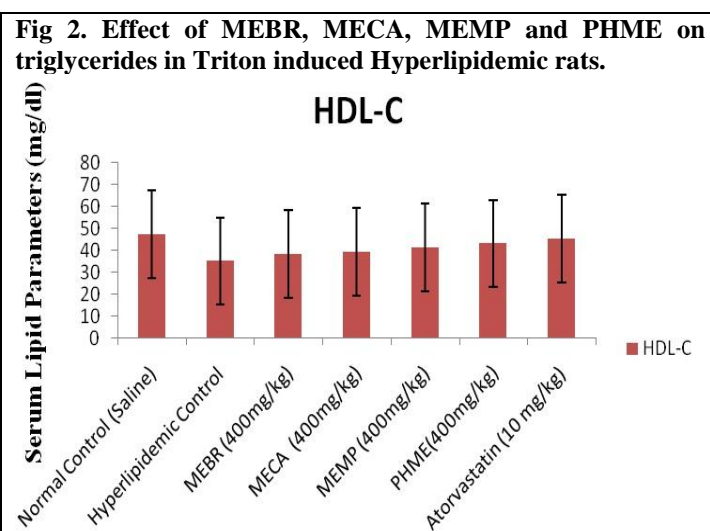
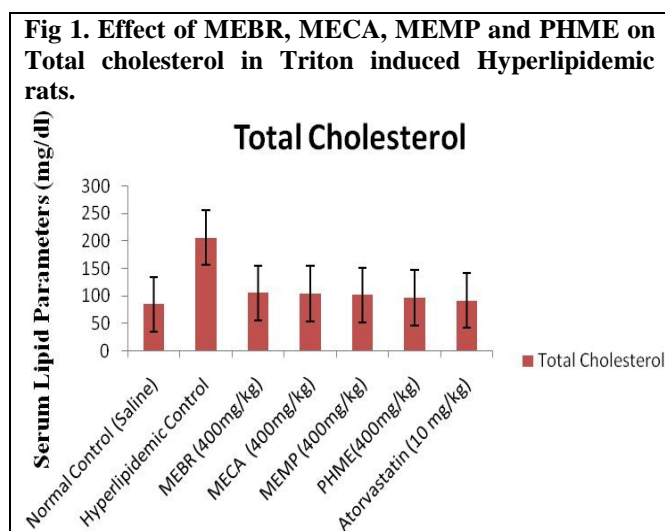
S. No	Groups	Serum Lipid Parameters (mg/dl)				
		Total Cholesterol	Triglycerides	HDL-C	LDL-C	VLDL-C
1	Normal Control (Saline)	84.67±1.18	64.73±7.07	47.27 ±3.62	24.46 ±1.61	12.95±1.71
2	Hyperlipidemic Control	205.7±13.81	117.9 ±5.45	34.98±4.40	147.1±16.1	23.58±1.39*
3	MEBR (400mg/kg)	105.5±11.22	104.2±7.11	38.23±2.31	43.02±5.12	21.11±1.44
4	MECA (400mg/kg)	104.1±11.34	103.1±5.1	39.22±3.12	42.11±4.42	20.13±1.22
5	MEMP (400mg/kg)	101.9±11.27**	99.12±2.56*	41.07±5.61*	41.01±3.62*	19.81±0.45*
6	PHME(400mg/kg)	96.36±14.16**	89.19±2.30*	43.03±3.66**	36.09±12.01*	17.83±0.46*
7	Atorvastatin (10 mg/kg)	91.17±12.21**	84.32±3.13**	45.10±2.69*	32.44±13.90*	16.86±0.70*

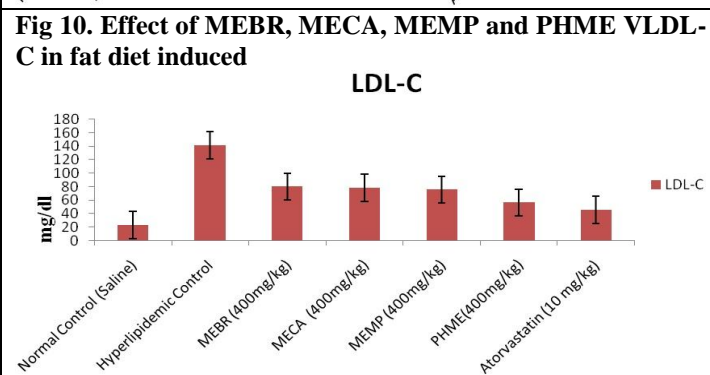
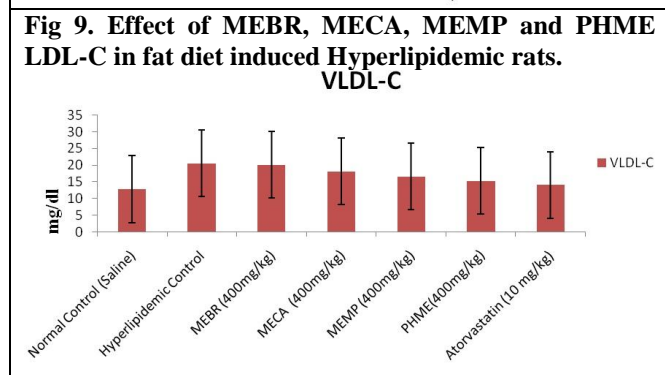
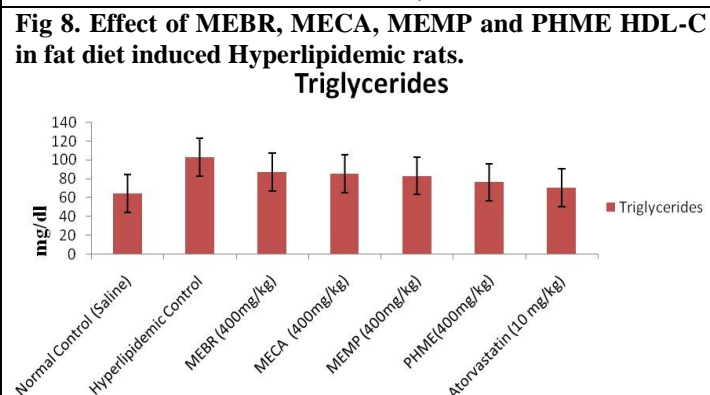
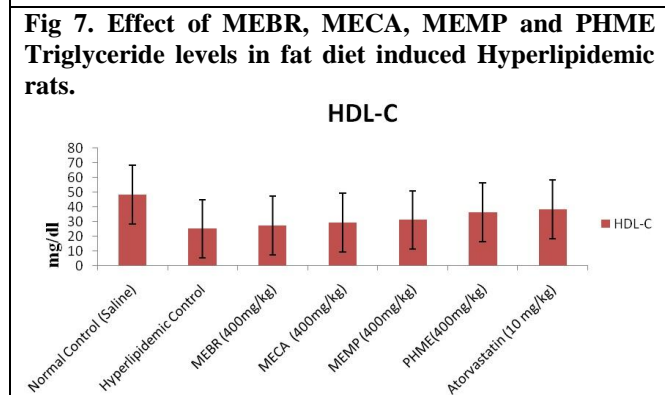
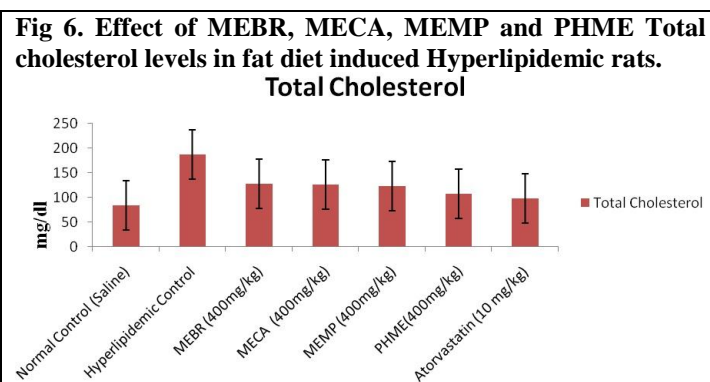
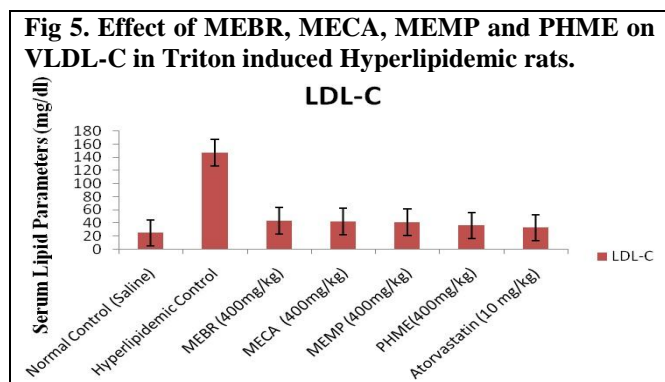
Values are mean ± SEM (n=6). Values are statistically significant at **P≤0.01 vs hyperlipidemic control using one way ANOVA followed by Dunnet’s test

Table 2. Effect of MEBR, MECA, MEMP and PHME serum lipid parameter levels in fat diet induced Hyperlipidemic rats

S. No	Groups	Serum Lipid Parameters (mg/dl)				
		Total Cholesterol	Triglycerides	HDL-C	LDL-C	VLDL-C
1	Normal Control (Saline)	83.84± 1.32	64.07 ± 7.13	48.34 ±5.58	22.69± 5.34	12.81± 1.27
2	Hyperlipidemic Control	187.0±11.85	102.9±5.28	25.05±3.43	141.4±13.04	20.58±1.24
3	MEBR (400mg/kg)	127.1±11.22*	87.11±3.21*	27.11±3.21*	80.11±12.12*	20.12±1.05*
4	MECA (400mg/kg)	125.2±11.41*	85.12±2.12*	29.11±2.21*	78.11±12.21*	18.11±1.09*
5	MEMP (400mg/kg)	123.0±11.23*	83.16±3.56*	31.00±4.36*	75.41±12.34*	16.59±0.81*
6	PHME (400mg/kg)	107.7±12.34**	76.28±5.66**	36.19±5.77**	56.25± 3.14*	15.23±1.25*
7	Atorvastatin (10 mg/kg)	97.62±11.64**	70.24±3.20*	38.34±2.4*	45.28±13.24*	14.00±0.47*

Values are mean ± SEM (n=6). Values are statistically significant at **P≤0.01 vs hyperlipidemic control using one way ANOVA followed by Dunnet’s test





CONCLUSION

Plants have played a significant role in human health care since the ancient times. Traditional plants exerts great role in discovery of new drugs, however, the exact mechanism responsible for activities is currently unclear. Therefore, further investigations need to be carried out to isolate and identify specific compounds present in the plant extract responsible for these activities and exact mechanism. It was concluded that individual extraction

shows the activities but in combination of plants it shows synergistic effect so poly herbal extraction is useful more when compared with given in individual plants.

ACKNOWLEDGEMENTS

I am grateful to the Department of Pharmacology, Vignan Institute of Pharmaceutical Sciences for providing research facilities for in vivo pharmacological studies in the present investigation.

REFERENCES

1. Robert W, Thomas P. The Pharmacological Basis of Therapeutics, 11th Edn, McGraw-Hill medical publication, 2006, 977-978.
2. Stone NJ, Robinson J, Lichtenstein AH, Merz NB, Blum CB. Guideline on the Treatment of Blood Cholesterol to Reduce Atherosclerotic Cardiovascular Risk in Adults: A Report of the American College of Cardiology/American Heart Association Task Force on Practice Guidelines. American heart association, 2013.
3. Kasper D L, Braunwald E, Fauci A S, Hauser, Longo, Jameson. Harrison's principles of internal medicine.16th Edn; Mc Graw Hill, 2008, 2288.

4. Rang HP, Dale MM, Ritter JM, and Flower RJ. Rang and Dale's Pharmacology, 6th Edn, 324.
5. Nicki RC, Brian RW, Stuart HR. Davidson's Principle and Practice of Medicine, 21st Edn, 2010, 577-579.
6. Walker R and Edward C. Clinical Pharmacy and Therapeutics, 3rd Edn, 2003, 353.
7. Mishra LC. Scientific Basis For Ayurvedic Therapies, CRC press LLC, 2004, 4
8. Nadkarni KN and Nadkarni AK. The Indian Materia Medica, Popular Prakashan, 1, 561.
9. Muhammad A and Sirat HM. Cox-2 inhibitors from stem bark of Bauhinia rufescens Lam. (Fabaceae). *EXCLI Journal*, 12, 2013, 824-830.
10. Anushia C, Sampathkumar P, Ramkumar L. Antibacterial and antioxidant activities of Cassia auriculata. *Global Journal of Pharmacology*, 3(3), 2009, 127-130.
11. Sutar NG, Sutar UN, Behera BC. Antidiabetic activity of the leaves of Mimosa pudica Linn. In albino rats. *Journal of Herbal Medicine and Toxicology*, 3(1), 2009, 123 – 126.
12. Dulloo A J, Seydoux L, Girardier P, Chantre V. Green tea and thermogenesis: interactions between catechin-polyphenols, caffeine and sympathetic activity. *Int. J. Obesity*, 2(252-8), 2000, 56.
13. Tokimitsu I. Effects of tea catechins on lipid metabolism and body fat accumulation. *Biofac*, 22(1-4), 2004, 141-3.
14. Ito Y T, Ichikawa Y, Morohoshi T, Nakamura S, Ishihara K. Effect of tea catechins on body fat accumulation in rats fed a normal diet. *Biomed Res*, 291, 2008, 27-32.